First report of *Psecas euoplus* Chamberlin & Ivie, 1936 from Colombia, with new salticid records for the department of Córdoba (Araneae, Salticidae)

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Abstract. *Psecas euoplus* Chamberlin & Ivie, 1936 is recorded for the first time from Colombia. In addition, the known distribution of the species *Lyssomanes amazonicus* G.W. Peckham, E.G. Peckham & Wheeler, 1889, *Lyssomanes remotus* G.W. Peckham & E.G. Peckham, 1896, and *Sarinda armata* (G.W. Peckham & E.G. Peckham, 1892), are extended to the department of Córdoba. We provide descriptions of the species reported here as well as images and drawings of both type material and our new records.

Key words. Colombian Caribbean, jumping spiders, Neotropics, taxonomy, zoogeography

Suárez-Martínez LA, Bedoya-Roqueme E, Quirós-Rodríguez JA (2024) First report of *Psecas euoplus* Chamberlin & Ivie, 1936 from Colombia, with new salticid records for the department of Córdoba (Araneae, Salticidae). Check List 20 (6): 1357–1371. https://doi.org/10.15560/20.6.1357

INTRODUCTION

The family Salticidae Blackwall, 1841, commonly known as jumping spiders, is the most species-rich family, including 688 genera and 6749 species (World Spider Catalog 2024). Currently, 156 species of salticid spiders, placed in 67 genera, are known from Colombia (Metzner 2024). In the Córdoba Department in the Colombian Caribbean Region, 33 species have been recorded, mainly from the low elevations of the northern region of the department (Bedoya-Roqueme and Lopez-Villada 2020; Suarez-Martinez and Bedoya-Roqueme 2021; Bedoya-Roqueme et al. 2022; Suarez-Martinez et al. 2022; Suárez-Martínez and Quirós-Rodríquez 2024).

Considering the number of habitats in the Department of Córdoba and the diversity of Neotropical Salticidae in general, species diversity is likely much higher than currently known (Bedoya-Roqueme and Lopez-Villada 2020). Due to its high diversity in species and habitats, this region is considered a high-priority zone with an estimated 60% chance of finding endemic species (Bedoya-Roqueme et al. 2022).

This study aims to report on several new records for both Colombia and the Department of Córdoba. For example, we record *Psecas euoplus* Chamberlin & Ivie, 1936 from Colombia for the first time. Additionally, we formally describe the female genitalia of *P. euoplus* and compare our male individuals to the allotype from Chickering's collection. This paper contributes significant data on the jumping spider fauna in Córdoba, the Colombian Caribbean as well as the country and region.

METHODS

The sampling of the jumping spiders was conducted during daylight hours (8:00–17:00 h), using (1) direct collection from shrubs and bushes (Coddington et al. 1991) and (2) beating of upper vegetation with beating sheets (Rubio et al. 2018). Collected spiders were placed in vials containing 70% alcohol. The epigyne of females was dissected following Levi (1965), treated with 10% lactic acid to digest soft tissue, and cleared in clove oil. Likewise, the pedipalp of males was dissected for observation in temporary mounts using 70% alcohol under a Carl Zeiss stereomicroscope. Multifocal photographs of the genitalia were taken using an



Academic editor: Tobias Bauer Received: 24 July 2024 Accepted: 14 November 2024 Published: 10 December 2024

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HD digital camera attached to a Carl Zeiss stereomicroscope and then merged with AxioVision Carl Zeiss SE64 (rel. 4.9.1. SP3). Measurements are in millimeters and taken using a micrometer connected to a Carl Zeiss stereomicroscope, Axiostar, in conjunction with AxioVision Carl Zeiss SE64 software (rel. 4.9.1. SP3). The identification of species was verified using the World Catalog of Spiders (2024) and the Jumping Spiders Catalog (Metzner 2024). Identifications were compared with the type material of each species deposited in the Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, United States (MCZ) (examined by photographs). Morphological terms follow Galiano (1963). The examined material has been deposited in the Entomology Laboratory of the University of Córdoba, Colombia (LEUC). Occurrence data were obtained through a review of scientific literature in the World Spider Catalog (2024), the Jumping Spiders Catalog (Metzner 2024), and the Global Biodiversity Facility (GBIF 2024). The maps were prepared using QGIS v. 3.32.0. (QGIS 2023).

The following abbreviations are used throughout. AERW = anterior eye row width; AL = abdomen length; CH = carapace height (~maximum); CL = carapace length; CW = carapace width; Em = embolus; F = Femur; LOQ = length of ocular quadrangle (ALE—PLE inclusive); M = metatarsus; MA = median apophysis; P = patella; PERW = posterior eye row width; PMEP = posterior median eye position (as ratio of ALE—PME distance to ALE—PLE distance); RTA = retrolateral tibial apophysis; ST = spermatheca; T = tibia; TL = total length.

RESULTS

Salticidae Blackwall, 1841

Psecas euoplus Chamberlin & Ivie, 1936

Figures 1–5

New records. COLOMBIA – CÓRDOBA • Momil, Vereda Mata de Caña; 09.2777, -075.6816; 34 m alt.; 28.V. 2022; L. Suárez Martínez leg.; 29 LEUC; OARA-213 • same locality; 1.X.2022; L. Suárez Martínez leg.; 1 3, LEUC; OARA-214.

Type material examined (allotype). PANAMA — COLÓN • Canal Zone, Fort Sherman; 01.VIII.1939; A.M. Chickering leg.; 1 ♂, MCZ-IZ:92460.

Identification. The female is only somatically described in the original description without a diagnosis; however, the epigyne was depicted (Chamberlin and Ivie 1936: pl. 8, fig. 67). We therefore provide a diagnosis for the females below.

Diagnosis. Females of *P. euoplus* can be differentiated from other species of the genus by an atrium located in the middle of epigyne and inseminations ducts that curve toward the posterior and then turn upward to connect to the ST (Figures 4A, B, 5D, E).

Similarly, males can be easily distinguished from other species by palps with short RTA short, stout, and blunt; tarsal bulb slightly swollen, with a rough, broad, blunt, and very chitinized portion, and a short, stout, and gently curved Em (Figures 3A–F; 5A–C).

Description. Cephalothorax with blackish upper part and sides of the thoracic part, turning reddish-brown on the upper part of the thorax (Figure 2A); covered with silver-gray scales (Figure 2A). Sternum labium dusky brownish yellow, sternum lighter in center, and labium with a narrow light tip, endites reddish brown, with lighter edge on mesal side (Figure 2B). Chelicerae, dark reddish brown, with two prolateral teeth (medial is very small), and seven retrolateral teeth. Abdomen deep-red dorsally, with four broads, evenly

Figure 1. Habitus, live specimen of *Psecas euoplus* Chamberlin & Ivie, 1936 from Colombia, Córdoba, Momil, Mata de Caña.





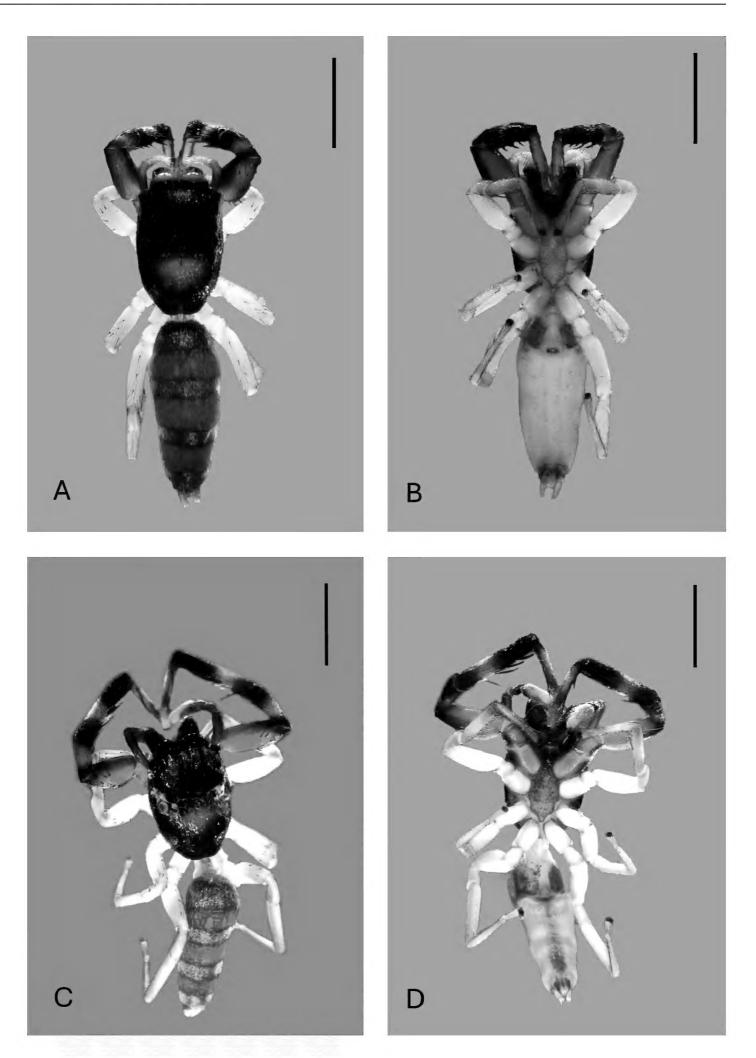


Figure 2. Habitus, *Psecas euoplus*Chamberlin & Ivie, 1936 from Colombia. **A.**Female, dorsal view. **B.** Female, ventral view. **C.** Male, dorsal view. **D.** Male, ventral view. Scale bars: 2.0 mm.

spaced transverse blackish bands, partially hidden by silvery-gray scales (Figures 2A). Habitus of the male as in the female (Figure 2C, D).

Epigyne with an atrium with two copular openings, located in the middle of epigyne, inseminations ducts that curve toward the posterior and then turn upward to connect to the ST (Figures 4A, B, 5D, E).

Palpus, femur curved ventrally in the basal side; RTA short, stout, and blunt; tarsal bulb slightly swollen, with rugulose, broad, blunt, highly chitinized side, and a short, stout, gently curved Em; cluster of five spines at tip (Figures 3A–F, 5A–C).

Measurements (mm). Females, TL = 6.94-7.22; CL = 2.65-2.89; AL = 3.86-4.25; CH = 1.02-1.19; CW = 1.84-1.94; AERW = 1.73-1.86; PERW = 1.61-1.72; LOQ = 1.27-1.35; PMEP = 0.41-0.42; eyes of the second row separated from the ALE by 0.37-0.39 mm; and from the PLE by 0.36-0.38 mm. Legs: F I: 1.39-1.71 II: 1.15-1.26, III: 1.23-1.32, IV: 1.75-1.88. P I: 1.08-1.14, II: 0.66-1.69, III: 1.60-0.64, IV: 1.77-1.78. Ti I: 1.52-1.73, II: 0.72-1.05, III 0.71-0.74, IV: 1.54-1.57 M I: 0.75-0.83, II: 0.66-0.72, III: 0.76-0.86, IV: 0.90-1.00. Ta I: 0.41-0.51, II: 0.47-0.50, III: 0.43-0.49, IV: 0.47-0.51. One male, TL = 6.61; CL = 2.95; AL = 3.33; CH = 1.23; CW = 1.80; AERW = 1.59; PERW = 1.07; LOQ = 1.18; PMEP = 0.41; eyes of the second row separated from the ALE by 0.37 mm and from the PLE by 0.37 mm. Legs: F I: 1.96 II: 1.10, III: 1.12, IV: 1.35. P I: 1.47, II: 0.74, III: 0.54, IV: 0.64. Ti I: 2.15, II: 1.16, III 0.70, IV: 1.43 M I: 1.35, II: 0.68, III: 0.73, IV: 1.09. Ta I: 0.57, II: 0.55, III: 0.49, IV: 0.41.

Figure 3. Psecas euoplus Chamberlin & Ivie, 1936: (A–C) male from Colombia, (D–E) male allotype (MCZ-IZ-92460).

A. Pedipalp, ventral view. B. Pedipalp, retrolateral view. C. Pedipalp, prolateral view. D. Male allotype (MCZ-IZ356 92460), ventral view. E. Male allotype (MCZ-IZ-92460), retrolateral view. F. Male allotype (MCZ-IZ-92460), prolateral view. The photographs were obtained by request to the Museum of Comparative Zoology, Harvard University (© President and Fellows of Harvard College). Scale

Figure 4. *Psecas euoplus* Chamberlin & Ivie, 1936 from Colombia. **A.** Epigyne, ventral view. **B.** Vulva, dorsal view. Scale

bars: 0.5 mm.

bars: 0.5 mm.

D В C В Α

Figure 5. Psecas euoplus from Colombia.

A. Pedipalp, ventral view. B. Pedipalp, retrolateral, view. C. Pedipalp, prolateral view. D. Epigyne, ventral view. E. Vulva,

dorsal view. Scale bars: A-C = 0.5 mm; D,

E = 0.2 mm.

Distribution. Known from Panamá, Barro Colorado Island (Chamberlin and Ivie 1936). Canal Zone, Biological Area; Porto Bello; El Valle, and Fort Sherman, (Chickering 1946). Darién: Cerro Pirre, head of Rio Setiganti (not published) (GBIF 2024). Brazil, Pará (not published) (GBIF 2024). Ecuador (Hill 2023). Colombia, vereda Mata de Caña, Momil, Córdoba (new record) (Figure 20D).

Comments. Specimens were collected on *Attalea butyracea*; (Mutis ex L.f.) Wess. Boer palm branches in a tropical dry forest fragment (Figure 21A).

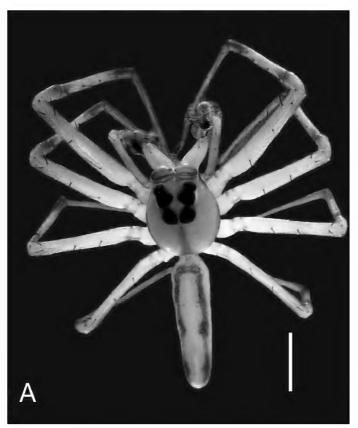
Lyssomanes amazonicus **G. W. Peckham, E.G Peckham & Wheeler, 1889** Figures 6–8

New record. COLOMBIA – CÓRDOBA • Momil, Vereda Mata de Caña; 09.2777, −075.6816; 28 m alt.; 28.V.2022; L. Suárez Martínez leg.; 2 ♂, LEUC; OARA-215.

Other material examined. COLOMBIA – META • Puerto López, Hacienda Mozambique; 01.1.1960; 1 ♂, MCZ-IZ-77533.

Identification. According to Galiano (1962), the males of *L. amazonicus* can be identified by the very irregular bulb, Em ensiform, very long and curved, the middle process, and the driver of the Em implanted on a large, and laminated formation (Figures 7A–C, 8A–C).

Distribution. known from Bolivia, Cochabamba: Rio Chapare, río Chimoré, (Platnick 1989; Galiano 1980). Brazil, Acre: Pimenteira, Xapurí, and Reserva Extrativista de Catuaba, Rio Branco. Amazonas: Manaus, Da Reserva Florestal Ducke; campus of the Universidade Federal do Amazonas (UFAM-Manaus); Igapó, Tarumã-Mirim, and Paraná do Xiboreninho. Pará, Teffé, Fonteboa (Peckham et. al. 1889; Petrunkevitch 1911; Galiano 1962; 1980; Logunov 2002; Bonaldo et al. 2009; Carvalho and Gasnier 2019); Pará: Belém, Universidade Federal do Pará, Mata Betina, and Cametá, Vila de Curuçambaba, Floresta (not published) (GBIF 2024). Colombia, Meta: Puerto López, Nariño: Tumaco, Bosque DIMAR, and Vereda Lagartera, Finca Maragricola (Logunov 2014; Galvis 2017). A new record from the Córdoba department. Ecuador, Orellana: Reserva Étnica Waorani (Logunov and Marusik 2003; Maddison 2015) Guyana, Bartica Dist: Kartabo; Mazaruni (Galiano 1980; Platnick 1989; Logunov 2014) (Figure 20A).



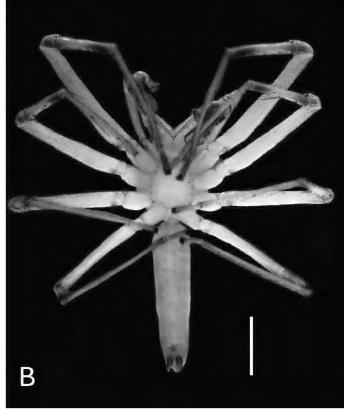
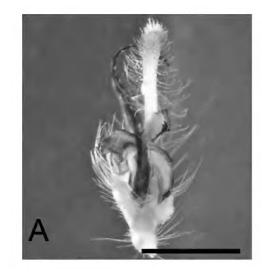


Figure 6. Habitus, *Lyssomanes amazonicus* G.W. Peckham, E.G. Peckham & Wheeler, 1889 from Momil, Mata de Caña. **A.** Male, dorsal view. **B.** Male, ventral view. Scale bars: 2.0 mm.

Figure 7. Lyssomanes amazonicus G.W. Peckham, E.G. Peckham & Wheeler, 1889 from Momil, Mata de Caña. **A.** Pedipalp, ventral view. **B.** Pedipalp, retrolateral, view. **C.** Pedipalp, prolateral view. Scale bars: 1.0 mm.







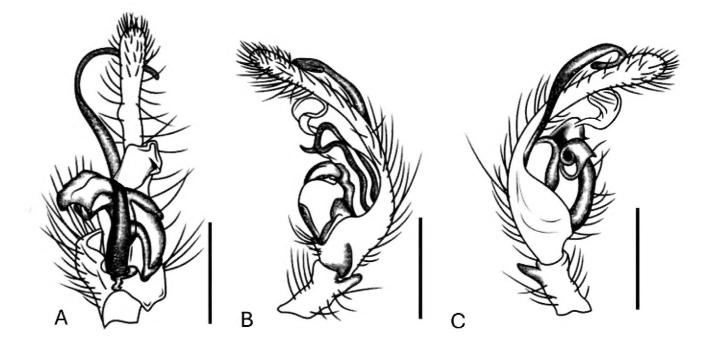


Figure 8. Lyssomanes amazonicus G.W. Peckham, E.G. Peckham & Wheeler, 1889 from Momil, Mata de Caña. **A.** Pedipalp, ventral view. **B.** Pedipalp, retrolateral, view. **C.** Pedipalp, prolateral view. Scale bars: 1.0 mm.

Lyssomanes bitaeniatus G.W. Peckham, E.G. Peckham & Wheeler, 1889 Figures 9–12

New records. COLOMBIA – CÓRDOBA • Momil, Vereda Mata de Caña; 09.2777, −075.6816; 31 m alt., 18.III.2022; L. Suárez Martínez leg.; 1 ♂, LEUC; OARA-216 • same locality; 31.XII.2022; L. Suárez Martínez leg.; 1 ♀, LEUC; OARA-217.

Other material examined. PANAMA — COLÓN • Canal Zone, Fort Sherman; 01.VIII.1939; A.M. Chickering leg.; 1 ♂, MCZ-IZ-91965. — PANAMA • Bella Vista; N. Banks leg.; 1 ♀, MCZ-IZ:91981.

Identification. According to Galiano (1962) the males of *L. bitaeniatus* can be identified by the very short, conical Em; on the retrolateral part of the bulb, the MA (short, black, and triangular) is observed, and a black chitinous lamina, curved inwards, between both, there is a longitudinal depression, at the bottom of this, the conductor of the Em (Figures 10A–F, 12A–C).

Distribution. Known from Brazil, Brasil, Ceará; Paraná; Pernambuco; Rio Grande do Sul (Mello-leitão 1943; Galiano 1980). Bahía: Paraguaçu; Minas Gerais: Belo Horizonte, Estação Ecológica da UFMG (not published). Colombia, Atlántico: Usiacurí, Reserva Campesina La Montaña; Bolívar: Cartagena, Isla Barú, and Turbaco, Jardín Botánico de Cartagena 'Guillermo Piñeres'; Cesar: Valledupar, Ecoparque Los Besotes, Monte Puma; Valle del Cauca: Cali, Buga; Rio Jamundí, entre Cali y Jamundí (Galiano 1980; Cabra-García et al. 2010; Galvis 2017); Magdalena: Bahía de Neguangue, Tayrona (not published) (GBIF 2024). Costa Rica, La Pacífica-Rio, and Upala (Logunov and Marusik 2003) Salvador, San Salvador, biological institute (Platnick 1989; Kraus 1955). Guatemala (Galiano 1980) Mexico, Chiapas: INIFAP Campus Experimental Rosario Izapa (not published) (GBIF 2024). Panama, Bella Vista; Canal Area, Fort Kobbe; Fort Sherman; Gamboa; Summit (Galiano 1980; Nentwig 1993; Logunov 2014) Venezuela, Caracas, San Sebastian (Peckham et al. 1889; Petrunkevitch 1911; Mello-Leitão 1943; Galiano 1962; 1980; Platnick 1989; Galiano 1980) (Figure 20B).

Figure 9. Habitus, *Lyssomanes* bitaeniatus G.W. Peckham, E.G. Peckham & Wheeler, 1889 from Momil, Mata de Caña. **A.** Male, dorsal view. **B.** Female, dorsal view Scale bars: 2.0 mm.



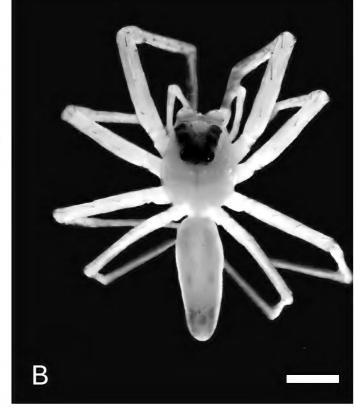


Figure 10. Lyssomanes bitaeniatus G.W. Peckham, E.G. Peckham & Wheeler, 1889: (A, C, E) male from Momil, Mata de Caña, (B, D, F) male (MCZ-360 IZ:91965).

A. Pedipalp, ventral view. B. Male (MCZ-IZ:91965), same view. C. Pedipalp, retrolateral view. D. Male (MCZ-360 IZ:91965), same view. E. Pedipalp, prolateral view. F. Male (MCZ-IZ:91965), same view. The photographs were obtained by request to the Museum of Comparative Zoology, Harvard University (© President and Fellows of Harvard College). Scale bars: 1.0 mm.

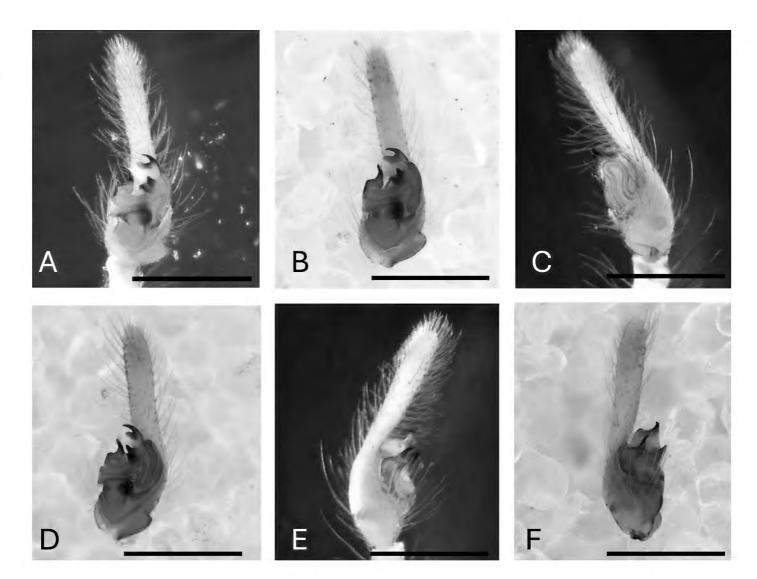
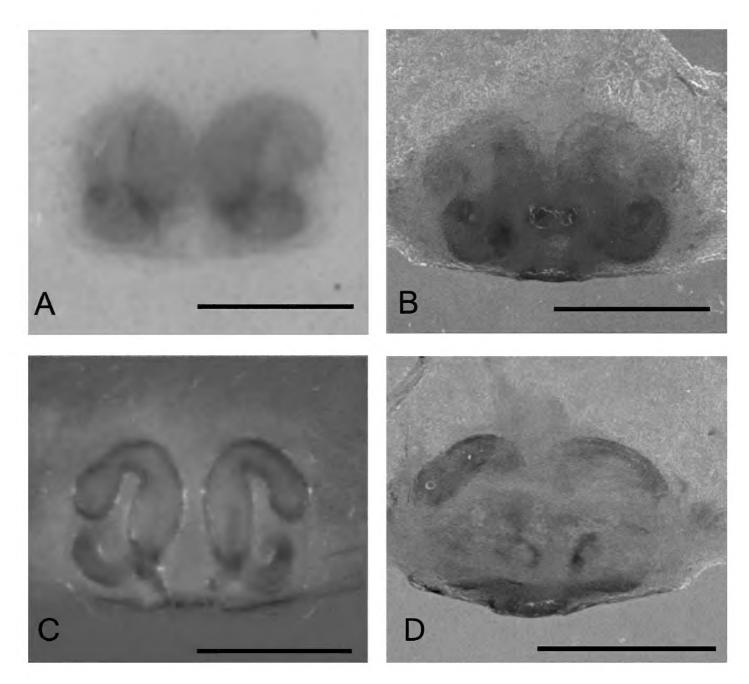


Figure 11. Lyssomanes bitaeniatus G.W. Peckham, E.G. Peckham & Wheeler, 1889: (A, C) female from Momil, Mata de Caña, (B, D) female (MCZ-IZ:91981). A. Epigyne, ventral view. B. Female (MCZ-IZ:91981), same view. C. Vulva, dorsal view. D. Female (MCZ-IZ:91981), same view. The photographs were obtained by request to the Museum of Comparative Zoology, Harvard University (© President and Fellows of Harvard College). Scale bars: 0.5 mm.



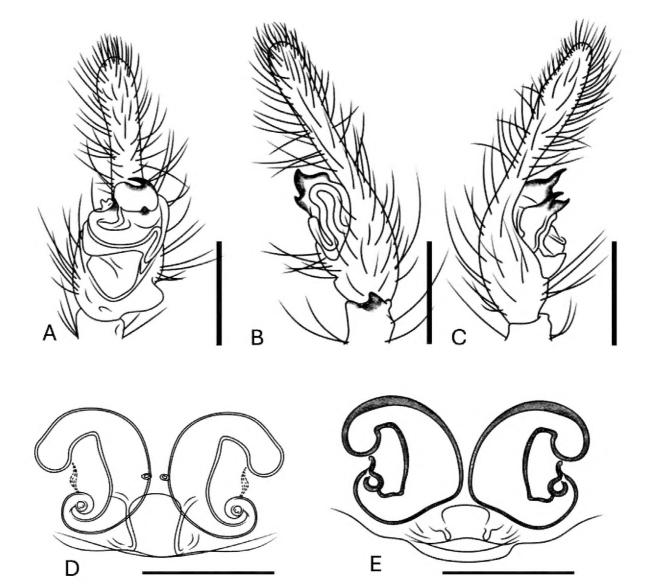


Figure 12. Lyssomanes bitaeniatus G.W. Peckham, E.G. Peckham & Wheeler, 1889 from Momil, Mata de Caña. **A.** Pedipalp, ventral view. **B.** Pedipalp, retrolateral, view. **C.** Pedipalp, prolateral view. **D.** Epigyne, ventral view. **E.** Vulva, dorsal view. Scale bars: A–C = 1.0 mm; D, E = 0.5 mm.

Lyssomanes remotus G.W. Peckham & E.G. Peckham, 1896

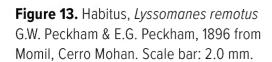
Figures 13-15

New record. COLOMBIA – CÓRDOBA • Momil, Cerro Mohan; 09.2232, -075.6634; 49 m alt.; 16.III.2022; L. Suárez Martínez leg.; 1 3, LEUC; OARA-218.

Type material examined (holotype). PANAMA − COCLÉ • Río Hato; 01.I.1850; E. Keyserling leg.; 1 ♂, MCZ-IZ:22883.

Identification. According to Galiano (1980) the males of *L. remotus* can be differentiated from the other species of the genus, by the long, divergent chelicerae; anterior side of the chelicerae with two internal angular spines and one external one, at the same height, an apical group of robust spines (8–10); three teeth on the promargin of the nail groove; six retrolateral teeth, two small ones located close together in the angle (large apical with a secondary tooth in its basal and internal part); and by inner spine of palpal bulb nearly (Figures 14A–F, 15A–C).

Distribution. Known from Brazil, Manaus: Ponta Negra; Bahía: São Francisco do Conde; Pará: Belém; Sergipe: Campus UFSE, São Cristovão (Galiano 1980; Platnick 1989; Logunov 2002). São Paulo: Primavera, Usina Hydroelectrical Sérgio Motta (not published) (Metzner, 2024). Colombia, Bolívar: Cartagena, Isla



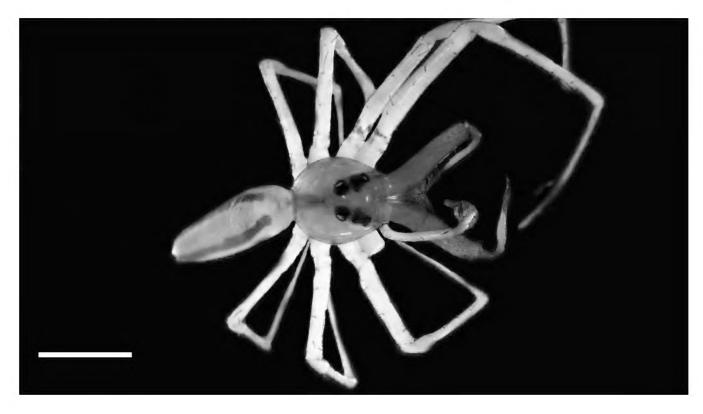
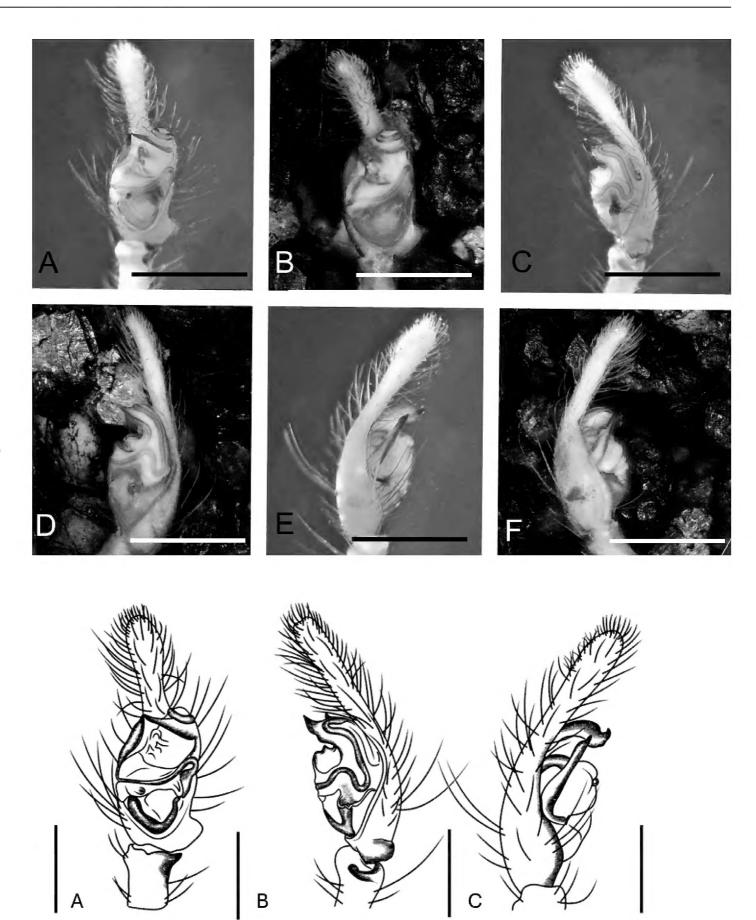


Figure 14. Lyssomanes remotus G.W.
Peckham & E.G. Peckham, 1896: (A, C, E) male from Momil, Cerro Mohan, (B, D, F) Male holotype (MCZ-IZ:22883).
A. Pedipalp, ventral view. B. Male holotype (MCZ-IZ:22883), same view.
C. Pedipalp, retrolateral view. D. Male holotype (MCZ-IZ:22883), same view.
E. Pedipalp, prolateral view. F. Male holotype (MCZ-IZ:22883), same view. The photographs were obtained by request to the Museum of Comparative Zoology, Harvard University (© President and Fellows of Harvard College). Scale bars: 1.0 mm.

Figure 15. *Lyssomanes remotus* G.W. Peckham & E.G. Peckham, 1896 from Momil, Cerro Mohan. **A.** Pedipalp, ventral view. **B.** Pedipalp, retrolateral, view. **C.** Pedipalp, prolateral view.



Barú, and Meta: Puerto López, Hacienda Mozambique (Logunov, 2014; Galvis 2017). A new record from the Córdoba department. French Guyana, Cayenne, Matauri (Logunov 2015). Guatemala (Pickard-Cambridge 1900; Petrunkevitch 1911; Galiano 1980). Panama, Gamboa (Platnick 1989; Pickard-Cambridge 1900; Galiano 1980; Peckham and Peckham 1896; Nentwig 1993). Trinidad and Tobago, Talparo River, Arena Forest (Cutler and Edwards 2002; Logunov 2014) (Figure 20C).

Sarinda armata (G.W. Peckham & E.G. Peckham, 1892)

Figures 16–19

New records. COLOMBIA – CÓRDOBA • Momil, Vereda Mata de Caña; 09.2777, −075.6816; 31 m alt.; 16.III.2022; L. Suárez Martínez leg.; 1 ♂, LEUC; OARA-222 • same locality; 18.III.2022; L. Suárez Martínez leg.; 1 ♀, LEUC; OARA-223.

Type material examined (syntype). PERU – LIMA • Lima; 01.1.1850; Peckham leg.; 1 ♂, MCZ-IZ: 20357.

Other material examined. PANAMA – COLÓN • Canal zone, Frijoles; 01.VIII.1936; A.M. Chickering leg.; 1♀, MCZ-IZ:23177 (holotype of *Sarinda strica* Chickering, 1946, junior synonym per Galiano (1965)).

Identification. According to Galiano (1965) the males of *S. armata* present a small apical retrolateral conical process in the tibia of the palp; disciforme bulb; Em short and thick, surround by the inferior and prolateral faces (Figures 17A–F, 19A–C). Females with epigyne weakly chitinized; four circular brown spots (the two lower ones are larger than the upper ones), separated by an oblique stripe; the duct makes two turns before emptying into the superior ST, to which it is linked by a spiral duct (Figures 18A–D, 19D, E).

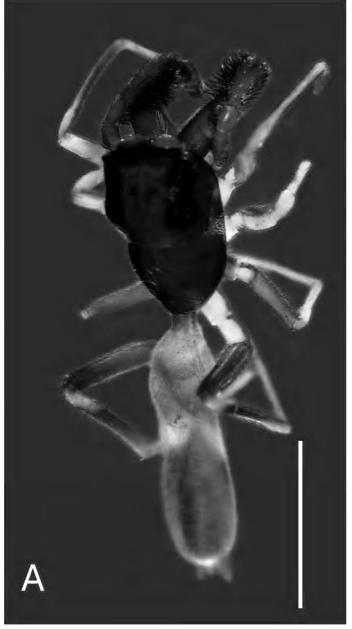
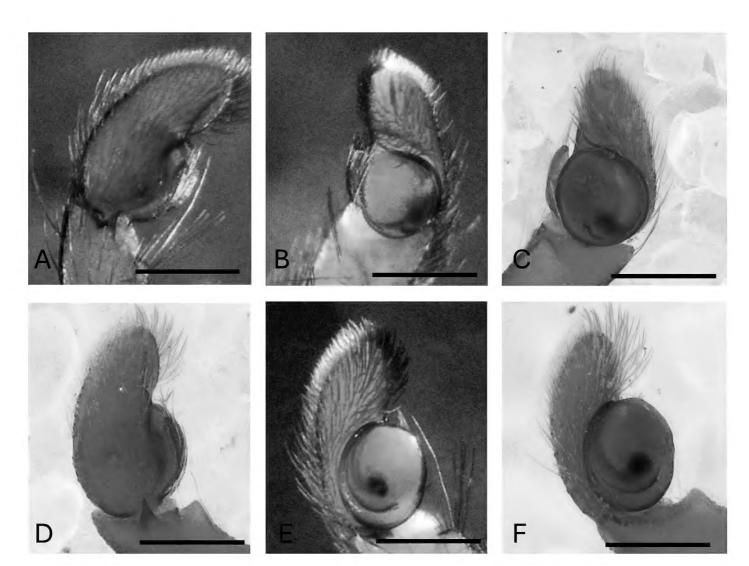




Figure 16. Habitus, *Sarinda armata* (G.W. Peckham & E.G. Peckham, 1892) from Momil, Mata de Caña. **A.** Male, dorsal view. **C.** Female, dorsal view. Scale bars: 2.0 mm.

Figure 17. Sarinda armata (G.W. Peckham & E G. Peckham, 1892): (A, B, E) male from Momil, Mata de Caña, (C, D, F) male syntype (MCZ-IZ:20357). A. Pedipalp, retrolateral view. B. Pedipalp, ventral view. C. Male syntype (MCZ-IZ:20357), same view. D. Male syntype (MCZ-IZ:20357), retrolateral view. E. Pedipalp, prolateral view. F. Male syntype (MCZ-IZ:20357), same view. The photographs were obtained by request to the Museum of Comparative Zoology, Harvard University (© President and Fellows of Harvard College). Scale bars: 0.5 mm.

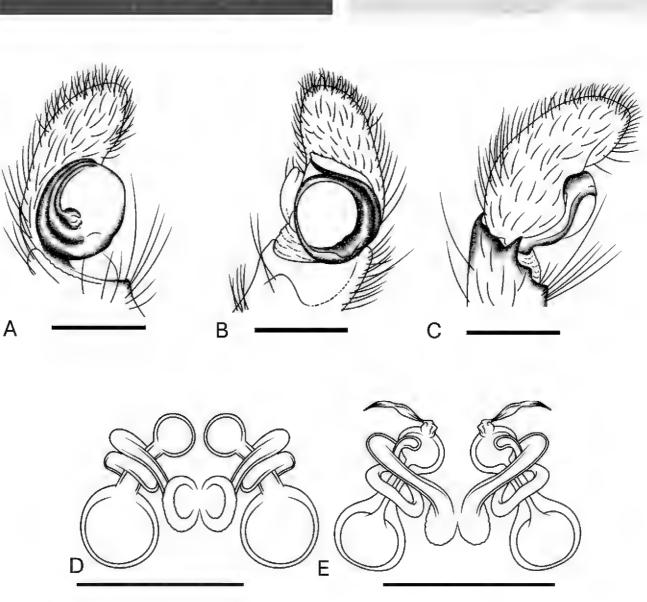


Distribution. Known from Colombia, Magdalena, Cañaverales, Tayrona Natural Park, and Rio Frio (Müller and Cutler 1989). A new record from the Córdoba department. Panama, Arraijan; Canal Zone; Biological Area; Bugaba; Isla Barro Colorado, and Porto Bello (Galiano 1965; Chickering 1946; Nentwig 1993; Platnick 1993). Perú, Lima (Peckham and Peckham 1892; Petrunkevitch 1911; Nentwig 1993; Platnick 1993) Trinidad. Venezuela (Petrunkevitch 1911) (Figure 20E).

Comment. The species was found on tree trunks and bushes (Figure 21A).

Figure 18. Sarinda armata (G.W. Peckham & E.G. Peckham, 1892): (A, C) female from Momil, Mata de Caña, (B, D) female (MCZ-IZ:23177). A. Epigyne, ventral view B. Female (MCZ-IZ:23177), same view. C. Vulva, dorsal view. D. Female (MCZ-IZ:23177), same view. The photographs were obtained by request to the Museum of Comparative Zoology, Harvard University (© President and Fellows of Harvard College). Scale bars: 0.5 mm.

Figure 19. Sarinda armata (G. W. Peckham & E. G. Peckham, 1892) from Momil, Mata de Caña. A. Pedipalp, prolateral view. B. Pedipalp, ventral, view. C. Pedipalp, retrolateral view. D. Epigyne, ventral view. E. Vulva, dorsal view. Scale bars: A–C = 0.5 mm; D, E = 0.2 mm.



DISCUSSION

Five species of salticids were collected. They were found in different microhabitats, mainly in areas of tropical dry forest in the subregion Bajo Sinú of the Colombian Caribbean. This increases the number of known salticid species in the department of Córdoba to 38 species. *P secas euoplus* is reported for the first time in Colombia, and the known distribution of the genus *Sarinda* Peckham & Peckham, 1892 is extended to include the department of Córdoba with with the finding of *S. armata*. Currently, two species of *Psecas* occur in Colombia: *P. jaguatirica* Mello-Leitão, 1941 in the south of the country and *P. rubrostriatus* Schmidt, 1956 in the Colombian Caribbean Region (Dupérré 2023; World Spider Catalog 2024). The

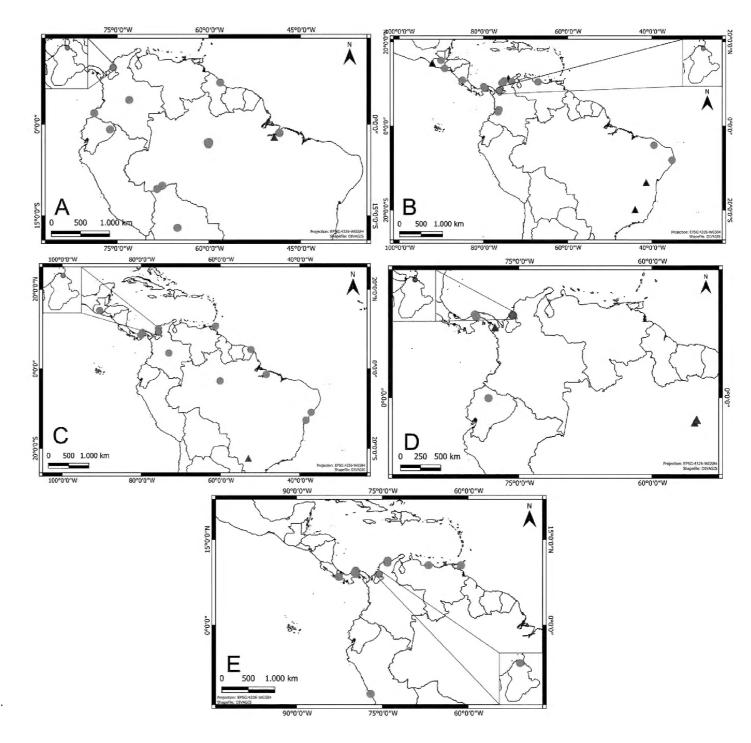
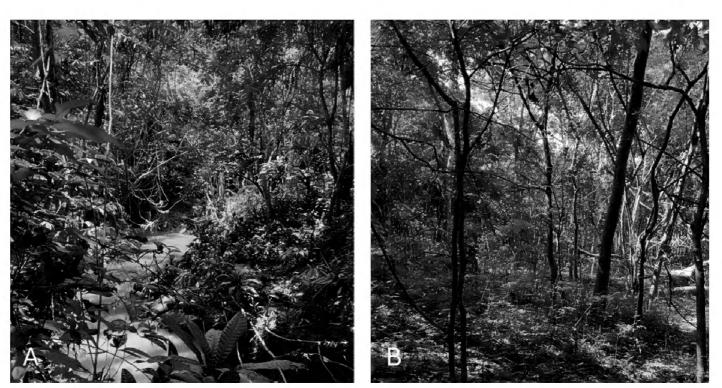


Figure 20. Known distribution of the jumping spiders recorded in this study. A. Lyssomanes amazonicus. B. Lyssomanes bitaeniatus. C. Lyssomanes remotus. D. Psecas euoplus. E. Sarinda armata. The blue circle indicates a new record for Colombia. The red circle indicates a new record for the department of Córdoba, Colombia. The yellow circle indicates previously published records. Black triangles indicate records from GBIF 2024.

Figure 21. Tropical dry forest in Córdoba, Colombia. **A.** Momil, vereda Mata de Caña. **B.** Momil, Cerro Mohán.



closest published record of *P. euoplus* to our report was made by Chickering (1946) in Canal Zone Biological Area (Panamá), which is located approximately 450 km away. Another record in GBIF (2024) from the MCZ, which was collected in 1961 and identified by M.E. Galiano, is from Darien, Cerro Pirre, head of Rio Setiganti (Panamá), about 268 km from the location of our new record. Therefore, our new record of this species represents a significant extension of the known geographic distribution and is the first report of it from South America.

Lyssomanes are a diverse group of jumping spiders widely distributed in the Americas, occurring from the United States of America to Argentina (Galvis 2020; World Spider Catalog 2024). Some species can mainly be found in the forest interior. In contrast, others prefer relatively open and sunny places, such as forest clearings or roadsides, and another group of Lyssomanes generally occurs on large, broad-leaved plants (Rubio et al. 2018; Galvis 2020). We collected L. amazonicus, L. bitaeniatus, and L. remotus from shrubs and tree branches in two fragments of little-disturbed tropical dry forest (Figure 21).

The genus *Sarinda* Peckham & Peckham, 1892 is distributed in the Americas, occurring from the United States of America to Argentina (World Spider Catalog 2024). In Colombia, this genus is currently represented by *S. atrata* (Taczanowski, 1871), *S. armata*, and *S. ruficeps* (Simon, 1901), which are all reported from San Marta, Colombian Caribbean (Müller and Cutler 1989). Here, we report *S. armata* from Momil, located in Córdoba, also in the Colombian Caribbean region.

This paper provides five new records for the department of Córdoba, including one new record for Colombia, considerably increasing the number of known salticid species in the region. However, most of the area and habitats of this Caribbean department, including urban and disturbed habitats, have not yet been sampled for salticids. Given the extremely high diversity of jumping spiders in South America (Metzner 2024), the 38 species now known very likely represent only a fraction of the full jumping spider diversity present in the department of Córdoba.

ACKNOWLEDGEMENTS

We thank Dr. Adam Baldinger and Dr. Ligia Rosario Benavides Silva of the Museum of Comparative Zoology (MCZ), Harvard University, for the collaboration in providing the photographs of type material, and to MCZ for permission to include these copyrighted images in our paper (© President and Fellows of Harvard College). We thank the members of the Study Group in Arachnology PALPATORES of the University of Cordoba, Colombia, for their assistance while sampling of the material. We give thanks to Biologist Carlos Andrés Nisperuza Pérez from the University of Córdoba, Montería, Colombia, for their collaboration and assistance in taking photographs. We thank Dr. María Florencia Nadal for her comments, advice, and assistance during the identification process. We thank reviewers (B.L. Pett and W. Maddison), as well as subject editor (Tobias Bauer) for their constructive comments on the manuscript. Finally, E. Bedoya-Roqueme thanks the agency Coordenação de Aperfeiçoamento de Pessoal de Nível Superior—CAPES for their graduation grant fellowship.

ADDITIONAL INFORMATION

Conflict of interest

The authors declare that no competing interests exist.

Ethical statement

No ethical statement is reported.

Funding

This study was financially supported by the vice-rectory of research and extension of the University of Córdoba.

Author contributions

Conceptualization: EBR, JQR, LSM. Investigation: LSM, EBR. Methodology: LSM, EBR. Resources: JQR. Project administration: JQR. Writing — original draft: EBR, JQR, LSM.

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Data availability

All data that support the findings of this study are available in the main text.

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